University of Rajasthan
Jaipur

SYLLABUS

M.Sc. Botany

(Semester Scheme)

III/IV Semester Examination 2018-19
1. Scheme of Examination:

1. Each theory paper EoSE shall carry 100 marks. The EoSE will be of 3 hours duration. Part A of theory paper shall contain 10 Short Answer Questions of 20 marks based on knowledge, understanding and applications of the topics/texts covered in the syllabus. Each question will carry two marks for correct answer.

2. Part “B” of paper will consist of four questions with internal choice (except in case where a different scheme is specified in the syllabus) of 20 marks each.

3. Each Laboratory EoSE will be of six hour duration and involve laboratory experiments/exercises, and viva-voce examination with weightage in ratio of 75:25.

2. Course Structure:

The details of the courses with code, title and the credits assigned are as given below.

Abbreviations Used

**Course Category**

CCC: Compulsory Core Course
ECC: Elective Core Course
OEC: Open Elective Course
SC: Supportive Course
SSC: Self Study Course
SEM: Seminar

Contact Hours
L: Lecture
T: Tutorial
P: Practical
S: Self Study

The medium of instruction and examination shall be English only.

Dy. Registrar
(Academic)
University of Rajasthan
JAIPUR
# M.Sc. BOTANY (SESSION 2018-19)
## THIRD SEMESTER

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Note:-

1. Elective core courses lab. can be opted only if the respective Elective theory has been opted by the student. ECC lab. Examination will be based on ECC lab. work of above papers wherever applicable (2 credits each)
2. Department will offer minimum three and maximum six theory elective courses for the semester based on options submitted by the students and availability of Faculty to teach the course.

**Theory Elective Courses:-**

**Theory Elective Clusters:-**

**Specialization Clusters:-**

A: PP: Plant Pathology
B: PM: Plant Morphology
C: PB: Plant Biosystematics
D: MI: Plant Microbiology
E: PPH: Plant Physiology
F: PE: Plant Ecology
G: GE: Plant Genetic Engineering
H: GEN: General

**Theory Elective Courses:-**

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<th>Course Title</th>
<th>Prerequisite</th>
<th>Semester In which course will be available</th>
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BOT 902- BIOSYSTEMATICS OF ANGIOSPERMS

Biosystemic studies: Population concept, methods of biosystematics, biosystematic categories - Ecads, Ecotypes, coenospecies, evolution and differentiation of species - various models. 4

International code of botanical nomenclature: principles, rules and recommendations. Taxonomic concept: Hierarchy, species, genus, family and other categories 10

Taxonomic literature: Flora, Monograph, Icones, Library, Manuals, Journals, Periodicals, Index. 4

Taxonomic tools and techniques: Herbarium: preparation, maintenance, utility, important national, international and digital herbaria, serological, Molecular technique, GIS and Mapping biodiversity. 10

Taxonomic evidences: Morphology, Anatomy, Palynology, Embryology, Cytology, Phytochemistry and Genome analysis. 8

Classification: Phenetic system; Phylogenetic- Cronquist, Dahlgren, Thorne and APG system with merits and demerits of these systems. 6

Salient features of the groups: Polypetalae, Gamopetalae, Monochlamydaceae and Monocotyledons. Study of the following families - Ranunculaceae, Caryophyllaceae, Sterculiaceae, Rhamnaceae, Leguminosae Cucurbitaceae, Apiaceae, Rubiaceae, Asteraceae, Asclepiadaceae, Apocynaceae, Convolvulaceae, Solanaceae, Acanthaceae, Lamiaceae, Polygonaceae, Chenopodiaceae, Amaranthaceae, Euphorbiaceae, Cyperaceae and Poaceae. 7

Phylogeny of Angiosperms: Ancestors of angiosperms with special reference to Amborella, time and place of origin of Angiosperms, Habit of Angiosperm, primitive living angiosperm, inter relationship among the major groups of Angiosperm. 6

Ethnobotany: Scope, Interdisciplinary approaches, ethnic groups of India, conservation practices of biodiversity, role of ethnobotany in national priorities, health care in India. 5

Suggested Readings:

Suggested Laboratory Exercises:

1. Description of a specimen from representative, locally available families.
   List of Locally Available Families
   (1) Ranunculaceae, (2) Capparidaceae, (3) Portulacaceae, (4) Caryophyllaceae, (5) Malvaceae,
   Convolvulaceae, (22) Solanaceae, (23) Boraginaceae, (24) Polemoniaceae, (25) Acanthaceae,
   Molluginaceae, (36) Euphorbiaceae, (37) Commelinaceae (38) Cyperaceae and (39) Poaceae

2. Description of a species based on various specimens to study intraspecific variation: a
   collective exercise.

3. Description of various species of a genus, location of key characters and preparation of keys
   at generic level.

4. Location of key characters and use of keys at family level.

5. Field trips within and around the campus, compilation of field notes and preparation of
   herbarium sheets of such plants, wild or cultivated, as are abundant.

6. Training in using floras and herbaria for identification of specimens described in the class

7. Demonstration of the utility of secondary metabolites in the taxonomy of some appropriate
   genera.

8. Comparison of different species of a genus and different genera of a family to calculate
   similarity coefficients and preparation of dendrograms.

9. Ethnobotany: Map of ethnic group in Rajasthan and India

Dy. Registrar
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BOT 903- MOLECULAR BIOLOGY

Genes and DNA
Genome, Gene, Evidences of DNA and RNA as the genetic material for bacteria, virus and eukaryotes. Double helical structure of DNA, Denaturation & Renaturation of DNA, DNA supercoiling. Coding DNA, ORF, Gene structure, Non-coding DNA & RNA.

DNA replication
Mechanisms of prokaryotic DNA replication: Initiation, Elongation and Termination; Replicons-linear, circular and D-loops; DNA polymerases, helicase, other enzymes and accessory proteins involved in DNA replication. Fidelity of replication and coordinating synthesis of the leading and lagging strands, Okazaki fragments.

DNA damage and repair: Causes of DNA damage and molecular mechanisms of repair - excision repair system in bacteria and eukaryotes, base excision, recombination repair systems and SOS repair.

Transcription and post transcriptional changes
Types of RNA, mRNA structure, prokaryotic and eukaryotic RNA polymerases, Transcriptional factors, promoter sequences, binding sites for RNA polymerase, transcription initiation, promoter clearance and elongation, termination, attenuation and antitermination. Role of Enhancers, repressors, mediators, silencers & inhibitors in transcription regulation.

RNA splicing and processing - capping, polyadenylation, splicing, spliciosome, mRNA stability, group I introns and transesterification, ribozymes and RNA editing.

Translation & protein targeting
Structure of tRNA, ribosome, Genetic Code, Translation-formation of initiation complex, initiation factors and their regulation, elongation and elongation factors, aminoacylation of tRNA, aminocyltRNA synthetase, and termination in Prokaryotes and Eukaryotes, translational inhibitors, Co and post translational modification of proteins.

Protein sorting and targeting of proteins into chloroplasts, mitochondria, vacuoles and peroxisomes Protein trafficking.

Gene regulation
Regulation of gene expression in prokaryotes: Basic models: Lac, arabinose and Tryp operons. Positive and negative controls. Regulation in viruses: Lytic and lysogenic cycle.

Techniques of gene expression analysis
Electrophoresis, blotting techniques (Southern, Northern and Western), gene sequencing methods (Sanger’s methods, Maxam Gilbert’s method, Automated and Pyrosequencing), nick translation, DNA fingerprinting, RNA protection assay, PCR (semi and quantitative) and Reverse Transcription-PCR, DNA microarrays, DNA-protein interaction (DNA footprinting). Protein-protein interaction (yeast two-hybrid system).

Suggested laboratory Exercises
1. Building of a model of B-DNA
2. Demonstration of linking number in supercoiled B-DNA.
3. Isolation of nuclei and identification of histones by SDS-PAGE.
4. Isolation of plant DNA and its quantitation by a spectrophotometric method.
5. Isolation of DNA and preparation of ‘cot’ curve.
7. Determination of size of DNA by comparison with DNA Ladder electrophoretically.
8. Determination of denaturation and renaturation of DNA double helix.
9. Isolation of RNA and quantitation by a spectrophotometric method.
10. Polymerase Chain reaction.
10. Immunological techniques: Ouchterlony method, ELISA and western blotting.

Suggested Readings:

See the following Review Journals:
Annual Review of Plant Physiology and Molecular Biology.
Current Advances in Plant Sciences.
Trends in Plant Sciences.
Nature Reviews: Molecular and Cell Biology.
Current Biology

BOT 904- Plant Physiology and Metabolism


Membrane transport: Passive non-mediated transport, Nernst equation, passive mediated transport. ATP- drive active transport, ion channels. Mineral nutrition: macro and micro nutrients and their role in plants

15

Biomolecules: General structure, classification, properties, distribution and functions of primary metabolites (carbohydrates, proteins, amino acids, lipids) and secondary metabolites (flavonoids, alkaloids, steroids)

Enzymes- Discovery and nomenclature, characteristics of enzymes. Concept of holzymes. apozymes, coenzyme, ribozyme, abzyme and artificial enzyme. Regulation of enzyme activity, mechanism of action. Michaelis-Menten equation


Nitrogen metabolism – N₂ fixation Symbiotic and Asymiotic. Nodule development Amino Acid Metabolism.

Fat metabolism: Synthesis of long chain fatty acids, lipid biosynthesis, & oxidation.
Plant growth regulators: Chemical nature, bioassay, physiological effects and mode of action of Auxins, Gibberelins, cytokinines, abscisic acid and ethylene, Salicylic Acid  
Strigolactones. Jasmonic Acid

Suggested Practical exercises

1. Effect of temperature and different solvents on membrane permeability of beet root.
2. Study of stomatal regulation.
4. Separation of chlorophyll by paper chromatography.
5. Separation of amino acids by paper chromatography.
7. Test of phenols, steroids, alkaloids and terpenoids.
8. Isolation of VAM fungi.
10. Separation of aliphatic wax components by thin layer chromatography (TLC).
11. Isolation of root nodule bacteria from roots of legume plant.
12. Quantification of protein in given plant sample.
14. Quantification of carbohydrates in given sample.
15. Demonstration of plasmolysis in *Rheo discolor* leaf peel.
17. The effect of pH on enzyme action.
18. To determine total soluble carbohydrates in given plant material.
19. To determine proline in given plant material.

Suggested Readings

BOT X01- PLANT REPRODUCTIVE BIOLOGY

Flower development and differentiation: Flower development, genetics of floral organ differentiation, homeotic mutants in Arabidopsis and Antirrhinum, sex determination.

Male gametophyte: Structure of anthers, microsporogenesis, role of tapetum, pollen development and gene expression, male sterility, sperm dimorphism and hybrid seed production, pollen germination, pollen tube growth and guidance, pollen storage, pollen allergy, pollen embryos

Female gametophyte: Ovule development, megasporogenesis, ultrastructural aspects of embryosac development.

Pollination, pollen-pistil interaction and fertilization: Floral characteristics in relation to pollination, structure of the pistil, pollen-pistil interactions, sporophytic and gametophytic self-incompatibility (cytological, biochemical and molecular aspects), double fertilization, in vitro fertilization.

Endosperm and Embryo development: Endosperm development, embryogenesis, storage proteins of endosperm and embryo, Polyembryony, apomixis, dynamics of fruit growth, biochemistry and molecular biology of fruit maturation. Embryo rescue in wide hybridization, Introduction to embryo culture, endosperm and ovary culture.

Latent life - Dormancy: importance and types of dormancy, seed dormancy, overcoming seed dormancy, bud dormancy.

Senescence and Programmed Cell Death (PCD): Basic concepts, types of cell death, PCD in the life cycle of plants, metabolic changes associated with senescence and its regulation, influence of hormones and environmental factors on senescence.

Suggested Readings


**Suggested practical/Field exercises:**

1. Study of microsporogenesis and gametogenesis in sections of anthers of different ages.

2. Examination of modes of anther dehiscence and collection of pollen grains for microscopic examination (maize, grasses, *Cannabis sativa*, *Crotalaria*, *Tradescantia*, *Brassica*, *Petunia*, *Solanum melongena*, etc.)


1. Pollen germination using hanging drop and sitting drop cultures, suspension culture and surface culture


7. Study of ovules in cleared preparations, study of monosporic, bisporic and tetrasporic types of embryo sac development through examination of permanent, stained serial sections.

8. Field study of several types of flower with different pollination mechanisms.

9. Emasculation, bagging and hand pollination to study pollen germination.

10. Study of nuclear and cellular endosperm through dissections and staining.

11. Isolation of zygotic globular, heart-shaped, torpedo stage and mature embryos from suitable seeds

12. Polyembryony in citrus, jamun (Syzygium cumini) etc. by dissections.

13. Endosperm, ovary and embryo culture.

14. Biochemical estimation (qualitative and quantitative) of metabolites of seeds.

Suggested Readings (for Laboratory Exercises)


**BOT X02- PLANT TISSUE CULTURE AND GENETIC TRANSFORMATION**

**Historical perspectives:** Concept of Cell culture, development of Tissue culture. Root tip culture, Embryo culture, Stem tip culture, Role of Auxin, Discovery of cytokinin, hormonal control of organ formation, Improvement of media, Preparation of & cloning of single cell culture. Contribution of various Scientists, Recent Advances

**Laboratory facilities, operation & Management:** Organization of laboratory, Equipments commonly used, media composition, preparation and handling, aseptic manipulation and preparation of explants.

**Cellular totipotency:** Process and mechanism, concept of differentiation, dedifferentiation & de-differentiation

**Cell culture and cell cloning:** Isolation of single cells, growth & subculture of suspension cultures, Types & culture medium for cell suspensions, synchronization & measurement of growth in suspensions, Viability of cultured cells, culture techniques and applications such as isolation and characterization of mutant plant cell cultures.
Virus free plants, Shoot tip culture, thermotherapy & meristem culture, Cryotherapy, Chemotherapy, Virus Indexing, Micropropagation: Proliferation of axillary buds, steps & advantages. Embryo culture and Embryo rescue for wide cross hybrids


**Organogenesis**: Direct and from callus & suspension cultures, Process and controlling factors.

**Anther culture** for Double Haploid Production; Androgenic and gynogenesis; Technique. Factors influencing anther culture, Pathways for development of Androgenic Haploids, Isolated Pollen culture method, genetic stability and variability of pollen-derived plants diploidisation of haploid plants, application of haploids in Plant Breeding.

Ovary and Endosperm culture.

**Somatic hybridization**: Isolation, Viability and culture of Protoplast, culture techniques. Protoplast fusion; types, methods, selection of somatic hybrids & cybrids by various methods. Somaclonal and gametoclonal variation and their selection: Causes of Variation, Induction of variation, detection and Isolation of variant, application in Plant breeding.

**Methods of plant transformation**: Direct gene transfer methods, Agrobacterium-mediated gene transfer, Biology of Agrobacterium, mechanisms of T-DNA transfer and integration. Chloroplast transformation and clean gene technology.

**Secondary metabolites**: Why do Plants Produce Secondary Metabolites? Process, methods and applications of Secondary Metabolites, Medium Composition and Effect of Nutrients, Elicitor-Induced Production of Secondary Metabolites, Effect of Environmental Factors, Biotransformation Using Plant Cell Cultures.

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**Microtechniques and analytical procedures used for Plant Cultures**: Histological techniques, TEM and SEM, Staining procedures for chromosome analysis, ELISA to certify pathogen free plants.

Role of Plant tissue culture in Biodiversity conservation and economic development. *In vitro* conservation method, conservation of endangered plant species, Cryopreservation for germplasm collection, Pollen and spore gene banks.

**Floriculture**: APEDA, Status in India, Economics, and Case studies: Dianthus

Tissue culture studies in medicinal plants, for rapid propagation of trees, for biomass Energy production, forestry, for-fruit trees and for improvement in cereal production.

**Agencies funding projects on Tissue culture**: CSIR, DBT, UGC, ICAR, DST & MoEF

Low cost options for tissue culture technology in developing countries: Integration of technology from lab to land, Green House features.

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**Suggested Readings:**


Bhojwani, Sant Saran, Dantu, Prem Kumar. Plant Tissue Culture: An Introductory Text


Suggested Laboratory Exercises:
1. Preparation of media for PTC.
2. Surface sterilization technique for preparation of explants.
3. Micro propagation technique
4. Organ culture.
5. Callus propagation, organogenesis, transfer of plants to soil.
6. Anther culture, production of Haploids.
7. Preparation of synthetic seeds
8. Cytological examination of regenerated plants.
9. Isolation of protoplasts from various plant tissues and testing their viability.
10. Single cell culture techniques: Bergmann plating technique, hanging drop & nurse cell culture.
11. Hairy root cultures
12. Elicitation and precursor feeding
13. Extraction of alkaloids and flavanoids from plant material and their separation using TLC.

BOT X03- Plant Ecology
Science of Ecology: Introduction to ecology, evolutionary ecology, ecological models:

Vegetation organization: Concepts of community and continuum, community coefficients, interspecific associations, ordination, species diversity and pattern diversity in community, concept of habitat and ecotone, ecological niche.
Vegetation development: Temporal changes (cyclic and non-cyclic), mechanism of ecological succession (relate floristic and initial floristic composition), succession models (facilitation, tolerance and inhibition models), Changes in ecosystem properties during succession, concept of climax.
systems: Nature and size of ecosystem, components of an ecosystem (producers, consumers and decomposers), Grazing (grassland) and Detritus food chain in freshwater ecosystems, food webs. Ecological energetic: Solar radiation and energy intakes at the earth's surface, energy flow models, Productivity of various ecosystems of the world and global biogeochemical cycles of carbon and nitrogen, Ecosystem Management: Homeostasis and cybernetics of ecosystem, Resilience of ecosystem, Restoration of degraded ecosystems, ecology of plant invasion. Ecosystem services,

Biomes, Biodiversity & Conservation: Major biomes of the world and Impact of changing climate on biomes, Biodiversity assessment (local, national and global), loss of diversity, conservation (ex-situ and in situ) and management, International Conservalional organizations, biodiversity act of India and related international conventions. Diversity indices, sustainable development, natural resource management in changing environment, molecular ecology, genetic analysis of single and multiple population, molecular approach to behavioural ecology, conservation genetics.

Suggested Readings

Suggested Laboratory Exercises

1. To determine minimum size and number of quadrat required for reliable estimate of biomass in grasslands.
2. To compare protected and unprotected grassland stands using community coefficients (similarity indices).
3. To estimate IVI of the species in a grassland/woodland using quadrat method.
4. To determine gross and net phytoplankton productivity by light and dark bottle method.
5. To determine soil moisture content, porosity and bulk density of soils collected from varying depths at different locations.
6. To determine the Water holding capacity of soils collected from different locations.
7. To determine percent organic carbon and organic matter in the soils of cropland, grassland and forest.
8. To estimate the dissolved oxygen content in eutrophic and oligotrophic water samples by azide modification of Wrinkler’s method.
9. To estimate chlorophyll content in $SO_2$ fumigated and unfumigated plants leaves.
10. To estimate rate of carbon dioxide evolution from different soils using soda lime or alkali absorption method.
11. To study environmental impact of a given developmental activity using checklist as a EIA method.

Theory Elective Courses
Specialization Clusters

Cluster: PP- Plant Pathology

BOT A01: PP: Plant Pathology-Principles and Diseases

History & Scope of Plant Pathology: Early developments and important landmarks in the history of plant pathology, Some Historical and Present Examples of Losses Caused by Plant Diseases 5

Dispersal of plant pathogens – Direct, Indirect and Biological Transmission 5

Host Factors in Disease Development: Inoculum Potential, Protective and defense mechanisms in plants, Phytoalexins, Genetics of plant disease, Phenomenon of resistance and susceptibility. Gene for gene hypothesis. Effect of pathogens on physiology of host plant. 15

Symptomology, Identification and Control of following plant diseases 15

Fungal Diseases.
Crucifers – Alternaria blight
Paddy – Paddy blast
Leaf rot and Foot rot of Pan (Piper beetle)
Red rot of Sugarcane, Flax rust,

Bacterial diseases
Brown rot of Potato,
Blight of Rice,
Crown Gall disease of stone fruits
Angular leaf spot of Cotton.

Phytoplasma Disease:
Little leaf of Brinjal,
Sesame Phyllody,

Plant Galls: Classification, developmental anatomy, host-parasite interaction and physiology of
Insect induced plant galls of Rajasthan. Economic significance of galls, Galls of Pongamia, Cordia, Prosopis and Salvador.

Nematology: General account of nematode disease, Symptoms caused by nematodes, Methods used in
Nematology: Control of plant parasitic nematodes, Inter relationship between nematode and other
Plant pathogens, Nematode Disease: Molya disease of Wheat and Barley. Soybean Cyst nematode

Elective Practical Lab BOT A11: Plant Pathology-Principles and Diseases

Suggested Laboratory Exercises:
Histopathology of nematode infected roots
Study of following diseases
Red rot of Sugarcane
Paddy Blast
Flax rust,
Crown Gall disease of stone fruits
Angular leaf spot of Cotton.
Little leaf of Brinjal,
Sesame Phyllody,
Galls of Pongamia, Cordia, Prosopis, Salvador
Molya disease of wheat and Barley
Soybean cyst nematodes
Any other practical based on theory.

Dy. Registrar
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JAIPUR
Recommended Books:


BOT A02: PP: Seed Science & Technology

History of seed testing and its importance to agriculture. Aims of seed testing. Seed-definition and its types. Sampling of seeds, purity analysis (physical and genetical), seed moisture content, germination test, rapid test of viability and evaluation, seedling evaluation, various methods of seed separation, cleaning, drying and Seed processing plant and its process.

Crops architecture of seed structure of angiosperms, identification and structure of seeds of important crop plants with special reference to Rajasthan (wheat, pearl millet, mustard, gram, p.m. spices). Identification of designated objectionable weeds at seed level.

Principles of seed production, seed production in self and cross pollinated crops; hybrid seed production. Production of foundation and certified seeds; synthetetic seed, terminator seed technology. Physiology of seed germination; seed and seedling vigour.

Seed storage methods, principles for safe seed storage, effects of storage, mycotoxins. Deterioration of seeds in storage by micro-organisms, insects and rodents; detoxification, control of seed deterioration, Seed.

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Elective Practical Lab BOT A12: Seed Science & Technology

Suggested Laboratory Exercises:

1. Structure of seeds of some crop plants (wheat, pearl millet, mustard, gram, and pea).
2. Preparation of inventory of designated objectionable weeds at seed level and identification.
3. Identification of seed coat cracking.
4. Study of physical purity of seed sample.
5. Study of seed germination, seedling abnormality and seedling index.
6. Determination of moisture content of seeds.
7. TZ test for seed viability.
10. Localization of starch, protein, lipids, tannins, phenols and lignin in seed sections.
11. Isolation and identification of storage fungi.
12. Preparation of phytosanitary certificate etc. of seed lot.

Suggested Readings:


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**BOT A03 : PP: Advanced Plant Pathology**

- **Plant Disease epidemiology and plant disease forecasting:** Computer simulation of epidemics, Methods used in Plant disease forecast, examples of Plant Disease forecasting System
  (10)

- **Disease Control:**
  - Immunizing the host
  - Disease control by transgenics
  - Innovative methods of plant disease control – Pollen Management and Integrated Pest Management
  (15)

- **Breeding for disease resistance**: Types of resistance, basal resistance, systemic resistance, acquired resistance, Gene for Gene concept, Production of disease resistant plants, Effectors.
  (15)

- **Molecular Plant Pathology:**
  Molecular diagnosis, Identification of genes and specific molecules in disease development, Genetics of host pathogen interaction, Molecular mechanism of resistance and Biotechnological approaches for disease
  (15)

- **Application of Biotechnology and Information technology in Plant Pathology & Integrated Pest Management.**
  (5)

**Elective Practical Lab BOT A13: Advanced Plant Pathology**

**Suggested Laboratory Exercises:**

- Biochemistry-Altered plant physiology due to plant pathogen interaction
- Histochemistry of altered metabolites-
  Protein, lipids, starch, cellulose, peroxidases and polyphenol oxidase
- Virus detection through biological (indicator hosts and host range) and serological methods (ELISA, Immunodiffusion)
- Virus Indexing
- Methods of application of fungicides – seed and foliar application
- Bio-control of plant pathogens – dual culture technique
- Bioassay of fungicides – poisoned food technique, inhibition zone technique and slide germination technique

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Recommended Books:


BOT A04: PP: Seed Pathology

Introduction and importance of Seed Pathology in modern agriculture. History of Seed Pathology. Various methods for testing seed borne fungi, bacteria and viruses (Dry seed examination, seed washing test, incubation methods, cultural, biochemical, serological, nucleic acid based methods).

Mechanism of seed infection and its types, environment influencing seed infection, extent of contaminated part of seed, morphology and anatomy of seeds in relation to invasion, stage of inoculum of the pathogen in seed- seed coat and pericarp, endosperm and胚 and embryo.

Seed borne diseases of some important crops with particular reference to the state of Rajasthan and India. Typical case of infection by: fungi (wheat- smuts and bunts, Sesame- basal rot), bacteria (Brassica- black rot, cluster bean- bacterial blight); viruses (tomato mosaic virus, pea seed borne mosaic virus,) and nematodes (wheat- ear cockle, rice- white tip).

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Seed-borne inoculum, inoculum density and assessment of seed borne inoculum in relation to plant infection, epiphytotics due to seed borne inoculum, disease forecast based on infected seed samples, tolerance limits of seed borne pathogens.

Transmission of seed borne disease: Systemic and non-systemic seed transmission, types of disease transmission, mode of establishment and course of disease from seed to seedling and plant, factors affecting seed transmission.

Management of seed-borne disease, principles of control, seed treatments (physical, chemical and biological), mechanism of action of seed treatments, major seed treatments for important seed borne pathogens and their methods of application.

Elective Practical Lab BOT A14: Seed Pathology

Suggested Laboratory Exercises:

1. Dry seed examination of seed lots.
2. Isolation and identification of seed-borne mycoflora by standard blotter method.
3. Preparation of culture media (PDA and NA).
4. Plating seeds on PDA/NA for identification of seed borne fungi and bacteria.
5. Other methods of plating e.g. deep freezing; 2,4D- blotter method.
7. Study of any seed borne nematode disease.
8. Detection of bacterial and viral pathogens in seeds.
9. LOPAT tests for detection of seed-borne bacteria.
10. Nucleic acid based detection of seed borne pathogens.
11. Histopathology of infected seed samples.
12. Physical control of seed-borne pathogens.
13. Antibiotic/ fungicidal assay against seed-borne pathogens
14. Biological control of seed-borne pathogens.
15. Field visits: Crop fields, FCI, NSC, Seed testing Labs., quarantine station (e.g. NBPG) etc.

Suggested Readings:


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Cluster: PM- Plant Morphology

BOT B01: PM: Angiosperm Morphology & Morphogenesis

- Floral anatomy and its role in explaining the morphology of the Stamen and Carpel, Seed and their taxonomic significance.
- Organizational relationship of anther tissues, ultrastructural aspects of microsporogenesis, sporoderm pattern. Pollen analysis, Pollen fertility and sterility, Allergy due to pollen.
- Storage and germination of pollen.
- Megasporogenesis - Basic types and their inter-relationships, ultrastructural aspects of pollen development. Pollen-pistil interaction, morphology of style and stigma, ultrastructural studies on pollen tube growth in the pistil, fertilization.

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Endosperm - Major types of endosperms and their development, endosperm haustoria, cytology of endosperm, role in embryo development.

Embryo - Embryogenic laws, Six major types of embryogenesis, development of Dicot and Monocot embryos, Apomixis- types and significance.

Embryological features of the following families: Santalaceae, Loranthaceae, Podostemaceae, Onagraceae, Cyperaceae, Exocarpaceae, Boraginaceae, Campanulaceae and Tropeolaceae.

Elective Practical Lab BOT B11: Angiosperm Morphology & Morphogenesis

Suggested Practical/Field Exercises:
1. Study of floral anatomy by cutting serial transverse sections.
2. Study different types of placentation.
3. Transverse sections of different stages of developing anthers to see development of wall layers and sporogenesis.
4. Spermoderm patterns of pollen grains
5. Make acetolysis preparations of pollen grains to study wall structure.
6. Pollen viability tests using different salt and sugar concentrations.
7. Endosperm haustoria
8. Stages in embryo development in plants like Raphanus and Legumes etc.

Suggested Readings
BOT B02: PM: Morphogenesis & Experimental Biology

Development and morphogenesis - Shoot apex the apical cell, meristem, the subcellular and biochemical struture of the meristem. the mechanism of primordium initaiton, transition to flowering, growth and formation of organs. Experimental work on shoot apical meristem, meristem culture and virus free plants, histochemical studies on apical meristems.

The phenomenon of morphogenesis - Correlation, polarity, symmetry, differentiation, regeneration.

Morphogenetic factors - Physical, mechanical, chemical and genetic factors. molecular basis of morphogenesis in plants with special reference to work done in Arabidopsis.


Microtechniques. Collection, killing and fixation (FAA and glutaraldehyde) of plant material, dehydration and embedding in paraffin and GMA, microtomy, (equipments and method), conventional and histochemical staining procedures. Transmission and scanning electron microscopy for internal structure and morphological development of plant organs and tissues.

Elective Practical Lab BOT B12: Morphogenesis & Experimental Biology

Suggested Practical/Field Exercises:
1. Study of organization of vegetative shoot apices
2. Study of organization of reproductive shoot apices.
3. Localisation of different metabolites through histochemical techniques.
4. Study of polarity of egg and synergids in embryo sac through permanent slides.
5. *In vitro* tissue and organ culture using different media. Preparation of media.
6. Inoculation and somatic embryogenesis.
7. Micropropagation and endosperm culture.
8. Histological techniques:
   - Collection, killing and fixation
   - Dehydration
   - Infiltration
   - Embedding
   - Microtomy
   - Staining
9. Microtoming equipments:
   - Microtome
   - Spreading table
   - oven
   - Ribbon box
   - Block holder
10. Demonstration of SEM and TEM
11. Micrometry

**Suggested Readings**

   Cambridge University Press, Cambridge

**Cluster: PB- Plant Biosystematics**

**BOT C01: PB: Advanced Biosystematics of Angiosperms**

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Aims, components, and principles of taxonomy: Alpha and omega taxonomy, documentation and scope. Concept of characters: analytic versus synthetic character, qualitative versus quantitative characters, good and bad characters.


Numerical Taxonomy- Principles, concepts, operational taxonomic units (OTU), data processing and taxonomic studies, taximetric methods for study of population variation and similarity - coding. Cluster analysis, cladistics and cladogram.

Suggested Readings:
Elective Practical Lab BOT C11: Advanced Biosystematics of Angiosperms

Suggested Laboratory Exercises:

1. Description of a specimen from representative, locally available families.

2. Description of a species based on various specimens to study intraspecific variation: a collective exercise.

3. Description of various species of a genus, location of key characters and preparation of keys at generic level.

4. Location of key characters and use of keys at family level.

5. Training in using floras and e-flora for identification of specimens described in the class.

6. Demonstration of the utility of secondary metabolites in the taxonomy of some appropriate genera.

7. Comparison of different species of a genus and different genera of a family to calculate similarity coefficients and preparation of dendrograms and cladograms.

8. Demonstration of the principle of priority.

9. Demonstration of the rejection of names according to ICBN.

10. Developed NPC formula and identified different species.
BOT C02: PB: Angiosperm Plant Biosystematics

Herbarium methods- Plant exploration, plant collection, pressing and drying, mounting, maintenance and importance of herbarium; Important national, international herbaria, concept of digital herbaria.  
Concept of species- speciation; Gradual and additive mechanism; species classification, concept of characters: analytic versus synthetic character, qualitative verses quantitative characters, good and bad characters. (10 Hours)  
Concept of population – its significance, types of variation (developmental, environmental and genetical), variance analysis, isolating mechanism.  
Ecotypes- Origin and differentiation, taxonomic significance of ecotypes, vicarians.  
Experimental taxonomy and hybridization- Role of hybridization in evolution, amphidiploidy, breeding barriers, epistasis and pleiotropy.  
Biochemical systematic- Methods and principles, systematic markers, PCR analysis, chemotaxonomy, seed proteins, technique of protein electrophoresis, chemical protein analysis procedures, genome analysis and nucleic acid hybridization.  

Suggested Readings:  

Elective Practical Lab BOT C12: Angiosperm Plant Biosystematics

Suggested Laboratory Exercises:
1. Flora writing
2. Synonymy
3. Taximetrics and cladistics
4. Molecular taxonomy

Cluster: MI- Plant Microbiology

BOT D01: MI: APPLIED PHYCOLOGY AND MYCOLOGY


Isolation, Purification and Culture of algae; Mass cultivation of microalgae with special reference to Spirulina and Dunaliella and their applications in human welfare. Cryopreservation, aquaculture (micro and macro algae cultivation).

Importance of algae in production of algal pigments, important bioactive molecules, role of algae in sustainable environment, role of algae in bioremediation, recent developments and future of algal biotechnology.

Applied Mycology: Application of fungi in food industry (Flavour & texture, Fermentation, Baking, Organic acids, Enzymes, Mycoproteins); Secondary metabolites (Pharmaceutical preparations); Fungi in agriculture (biofertilizer) and remediation of contaminated soils. Mycotoxins; Fungal endophytes of plants and their applications: Endophytic fungi, colonization and adaptation of endophytes. Endophytes as latent pathogens and biocontrol agents.

Fungi as Biological control agents (Mycofungicides, Mycoherbicides, Mycoinsecticides, Mycoonematicides). Fungi in plant disease control; Selection, production and formulation of

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Mushroom cultivation - General account of Oyster, white button, paddy straw, Morels, Truffles & Poisonous mushrooms. Cultivation of mushrooms (Agaricus bisporus, Pleurotus and Volvoriella), Medicinal and nutritional value of edible mushrooms, Effect of environmental, nutritional and chemical factors on mushroom cultivation (intensive and extensive cultivation methods).

Suggested Readings:

1. Kumar, H.D. Introductory Phycology; Affiliated East West Pvt. Ltd., New Delhi
7. Powel, C and D. J. Bagyaraj - V.A. Mycorrhizae

Elective Practical Lab BOT D11: Applied Phycology and Mycology

Suggested Laboratory Exercises:

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1. Isolation and culture of algae.
2. Identification of Algal biofertilizers.
3. Identification of toxic algae.
4. Identification of bloom forming algae.
5. Making an algal bloom.
6. Phytoplankton identification from local water bodies.
7. Isolation, culture and identification of fungi using moistened blotters, PDA and Sabouraud's Dextrose Agar media
8. Cultivation of *Spirulina* and *Dunaliea*.
10. Study of Mushroom specimens
11. Mushroom cultivation.
12. Demonstration of antagonistic fungi a) Antibiosis b) Competition c) Mycoparasitism
13. Mycorrhizae: ectomycorrhiza and endomycorrhiza (Photographs)
14. Study of Mycorrhizal colonization in roots of *Parthenium* and *Tagetes*.

**BOT D02: MI: APPLIED MICROBIOLOGY**

*Food and Dairy Microbiology*: Microbial spoilage of food products including cereals, fruits, vegetables, meat, fish, and dairy products, Factors influencing microbial growth in foods - extrinsic and intrinsic.

*Principles of food preservation*, Food preservatives and their uses, Fermented food, wine, dairy products, cereals, and milk products, Bacteriocins and their application in food preservative (Nisin, Lactococcuslactis), food additives. Nutritional value of fermented biological examination of milk and milk products, source of their contamination and control.

*Microorganisms in Waste Treatment*: Solid waste treatment (Landfills, incineration, composting, anaerobic digestion and pyrolysis). Waste water treatment: Pretreatment, primary, secondary (activated sludge, surface aeration basins, fluidised bed reactors, trickling filter, bioreactor, biological contactors, membrane bioreactors and secondary sedimentation) and tertiary treatment, disinfection and odor control; Application of biofilms in waste water treatment. Organisms as indicators of water quality.

Bioremediation: In situ & Ex situ remediation, Concept of bioremediation technologies, Microbial consortium, Microbial remediation of oil spills, paper and pulp mill effluents and textile effluents; Biostimulation and Bioaugmentation. Bioaccumulation of metals and detoxification. Genetically Modified Organisms released and its environmental impact assessment; Molecular approach to environmental management, Degradative plasmids, Genetic exchange in xenobiotic chemicals.


Mycoorrhizal fungi: Diversity of endo and ectomycorrhizal fungi, culturing and benefits, role in bioremediation of soil. Fungal endophytes as biocontrol agents. Agriculturally important fungi in sustainable agriculture with special emphasis on Biopesticides, Mycocideicides and Mycoinsecticides.

Suggested Readings:-

2. Singh, B.D. Biotechnology, Kalyani Publishers, New Delhi
13. Adams M.R and Moss M.O: Food microbiology: Royal society of chemistry


Elective Practical Lab BOT D12: Applied Microbiology

Suggested Laboratory Exercises:

1. Microbiological examination of food.
2. Detection of number of bacteria in milk by standard plate count (SPC).
4. Adulteration tests for milk.
5. Isolation and identification of Lactobacillus from fermented dairy products.
6. Isolation and identification of microorganisms from contaminated food and dairy samples.
7. Sampling and analysis of microbial load on food contact surfaces.
8. Microorganisms degrading oil/textile dyes/petrol.
11. Design and operation of multistage reactor for degradation of waste water.
12. Isolation of xenobiotic compound degrading bacteria by enrichment culture technique.

BOT D03: MI: MICROBIAL TECHNOLOGY

Prerequisite of industrial microorganisms; Strategies for screening, selection and improvement of industrial strains; Methods of preservation & maintenance of microbial & their stability; Formulation of fermentation media; methods of sterilization; operating techniques of microbial strains; inoculum preparation and inoculum development. View of upstream and Downstream processing, In situ recovery of products.

Microbial Fermentations: Metabolic pathways and metabolic control mechanisms, bacterial production of citric acid, lactic acid, enzymes (alpha-amylase, lipase, xylase, esterases, proteases), acetone-butanol, Amino acid (lysine and glutamic acid), Vitamins (flavin, cyanocobalamine).

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**Microbial Production of Therapeutic and Pharmaceutical Compounds:** Microbial production of therapeutic compounds (β lactam, aminoglycosides, Ansamycins (Rifamycin), peptide antibiotics Quinolones), biotransformation of steroids, Fermentative production of antibiotics (penicillins, erythromycins); Vaccines, recombinant vaccines.

**Biofuels:** Useful features of bio-fuels, The substrate digester and the microorganisms in the process of biogas production (biomethanation), Production of bioethanol from sugar, molasses, starch and cellulosic materials. Ethanol recovery, Microbial production of hydrogen gas, biodiesel from hydrocarbons.

**Modern trends in Microbial Production:** Microbial production of bioplastics (PHB, PHA), bioinsecticides (thiaciide), biopolymer (dextran, alginate, xanthan, pullulan), Biofertilizers (Rhizobia, BGA, Azotobacter, Phosphate solubilizing microorganisms), Single Cell Protein and production of biological weapons with reference to anthrax.

**Advances in Microbial Technology:** Recombinant fermentations, strategies for fermentation with recombinant organisms and stability issues of recombinants. Applications of immobilized/co-immobilized cells/enzymes in fermentation industry; Overview of nanomaterials and biosynthesis of silver and gold nanomaterials from microbes.

**Elective Practical Lab BOT D13: Microbial Technology**

**Suggested Laboratory Exercise:**

1. Screening of industrial microbes
2. Collection and identification of important bacterial/fungal strains of industrial importance
3. Determination of specific cell growth rate.
4. Production and characterization of citric acid using A. niger.
5. Microbial production of glutamic acid.
6. Production of rifamycin using Nocardia strain.
7. Comparison of ethanol production using various Organic wastes /raw Material [Free cells/ immobilized cells].
8. Laboratory scale production of Biofertilizers [Nitrogen fixer/Phosphate Solubilizers].
9. Microbial production of dextran by Leuconostoc mesenteroides
10. Microbial production of hydrogen gas by algae/bacteria
11. Laboratory production of vinegar.
12. Demonstration & operation of ultrasonicator
13. Enzymatic production of high fructose syrup from inulin
14. Production of alkaline phosphatase in lab scale Fermenter
16. Synthesis of nanoparticles using microbes
Recommended Readings:

13. Pharmaceutical Microbiology SS Purohit, AK Saluja and HN Kakrani, 2012

Cluster: PPH- Plant Physiology

BOT E01: PPH: Plant Biochemistry and Metabolism

Energy: Principles of thermodynamics, free energy, Concept of energy rich compounds and intermediates, Structure and function of ATP, ATP synthesis and chemo-osmotic hypothesis of ATP generation, Redox reaction, Types and mechanisms of Phosphorylation.

Nucleotides metabolism: Biosynthesis of Ribonucleotides and of Deoxy-ribonucleotides-salvage and de novo pathways, nucleotide degradation.

Enzymes: Enzyme kinetics, Michalis- Menten equation and significance of Km value, negative and positive co- operativity, enzyme nomenclature and EC number, Catalytic mechanisms: Acid-Base catalysis, covalent catalysis, metal ion catalysis, electrostatic state bonding, Lysozyme as model enzyme for catalytic mechanism, Regulation of enzyme activity: feedback and allosteric regulation, active sites, isozymes, activators and inhibitors.
Proteins: Structure, Ramachandran Plot, techniques of protein purification. Protein sequencing and proteomics, Dynamics of protein structure, Protein structure prediction tools, Protein Folding, protein stability, globular proteins and maintenance of specific confirmation, structural evolution. Molecular Chaperons, Protein Data Banks.

Secondary Metabolites: Detailed account of Coumarins, Lignins, Insecticides(pyrethrins and rotenoides), Tannins, Flavonoids, Alkaloids and Steroid and their role in plant defense.

Vitamins: Water and fat soluble vitamins, biochemical functions of thiamine, riboflavin, nicotinic acid, pentothenic acid, pyridoxine, biotin, folic acid, vitamin B12, ascorbic acid, Vitamin A and Vitamin D.

Suggested Readings

4. WilliamG.Hopkins and Norman P.Huner. (2009),Introduction to Plant Physiology,Published by John Wiley &Sons,Inc,USA.

Elective Practical Lab BOT E11: Plant Biochemistry and Metabolism

Suggested Laboratory Exercises:

1. Estimation of lipids
2. Separation of chlorophyll by paper chromatography.
5. Test of phenols, steroids, alkaloids and terpenoids.
6. Isolation of VAM fungi

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7. Quantification of protein in given plant sample.
8. Separation of aliphatic wax components by thin layer chromatography (TLC)
9. Quantification of carbohydrates in given sample.
10. Study of stomatal regulation.
11. Demonstration of Physiological effects of growth regulator.
12. Effect of temperature and different solvents on membrane permeability of beet root.
13. To determine proline in given plant material.
15. The effect of pH on enzyme action.
16. Determination of amylase activity
17. Demonstration of plasmolysis in *Rheo discolor* leaf peel.
18. Isolation of root nodule bacteria from roots of legume plant.

**BOT E02: PPH: STRESS BIOLOGY**

*Environmental stresses*: Introduction, significance, types

*Water deficit stress*: Effects on physiological processes, Physiological responses to water deficit stress, Various strategies of drought resistance in plants, Mechanism of stomatal action, ABA as stress hormone, ABA dependent and ABA independent pathways, LEA proteins

*Flooding stress (anoxia)*: Nature of waterlogging stress, Effects of flooding stress on physiological processes in plants, Wetland and non-wetland species, Mechanism of waterlogging tolerance

*Salinity stress*: Definition of saline soil, Causes of soil salinization, A brief outline of salt affected soils in India, Salinity stress- combination of osmotic stress and ionic stress, Physiological responses of plants to salinity stress, mechanism of salinity tolerance in higher plants, Signaling under salinity stress- Salt Overly Sensitive (SOS) Pathway, Compatible osmolytes, role in osmotic adjustment

*Heavy metal stress*: Effect of ion toxicity (iron, zinc), heavy metals toxicity and aluminum toxicity in plants, Mechanism of aluminum tolerance, Phytoremediation

*Thermal stresses*: Effects of high and low temperatures on plant, Physiological responses of plants to high and low temperatures, Mechanisms of high and low temperatures tolerance, Role of Heat Shock Proteins (HSPs), calcium, calmodulin and C-repeat binding factors (CBFs)

*Oxidative stress*: Influence of high light intensity on photosynthesis, Generation of reactive oxygen species (ROS), Effects of ROS, Photoprotection and ROS detoxification mechanisms
plants

**Biotic stresses**: Responses of plants towards biotic stresses, plant defense system, genetic basis, understanding of R genes, Systemic acquired resistance

**Biotic stress signaling (plant defense)**: Elicitors and plant defense: Plant responses to elicitors and their role in crop improvement, Plant defense priming, Role of Jasmonic acid, salicylic acid, ethylene and nitric oxide signaling in plant defense.

**Suggested Readings**:

2. Biochemistry and molecular biology of plants: Buchanan et al 2000, American Society of Plant Biologists, USA
4. Plant Responses to Abiotic Stress: Hirt and Shinozaki: (Online) Springer
5. Plant Responses to environmental stress: Smallwood et al BIOS Scientific Publishers
8. Plant Ecophysiology: Prasad MNV 1997, John Wiley & Sons, Inc, USA

**Elective Practical Lab BOT E12: Stress Biology**

**Suggested Laboratory Exercises**:

2. Leaf disc assays under various abiotic stresses.
3. Determination of chlorophyll content under stress.
5. Estimation of sodium, potassium, chlorides in different plant leaves.
6. Estimation of free amino acids content in the given sample
7. Study of effect of fungal infection on peroxidase activity.
8. Study of phenolics in scales of onion varieties differing in disease resistance.
10. Estimation of proline content in given water stressed samples
11. Study of seed germination under stress condition.

12. Estimation of betaine content in salt stressed samples.

13. Determination of ascorbic content in temperature (low and high) and salt stressed samples.

14. Determination of the concentration of polyamines in the given stressed samples.

15. Effect of stress on the activity of following scavenging enzymes:
   (a) superoxide dismutase, (b) catalase, (c) peroxidase, (d) ascorbate peroxidase.


17. Effect of stress on membrane damage in relation to lipid peroxidation.

18. Effect of water stress and Hyperthermia on the activity of nitrate reductase.

**BOTE03: PPH: ADVANCED PLANT PHYSIOLOGY**

*Signal transduction in plants:* Receptors and G-Proteins, secondary messengers, phospholipid signaling, role of cyclic nucleotides, calcium-calmodulin cascade, diversity of protein kinases and phosphate, single transduction mechanism with special reference to pathways activated by Auxins, Gibberellins, Cytokinins, ethylene and Abscisic acid

**Photomorphogenesis: Responding to light:** Photoreceptors, Phytochrome: Responding to Red and Far-Red Light, Cryptochrome: Responding to Blue and UV-Light. Phytochrome and Cryptochrome mediate numerous Developmental Responses - Seed germination, De-Etiolation, Shade avoidance, Control of Anthocyanin Biosynthesis, Rapid phytochrome response. Chemistry and mode of action of Phytochrome and Cryptochrome. Physiology of light induced responses - Tropic (Phototropism & Gravitropism) and Nastic movements; Stomatal movement; Molecular mechanism of actions of photomorphogenic receptors.

**Biological Clocks in plants:** Clock-driven Rythms persists under constant conditions, Role of light and temperature, Characteristics, mechanism and Significance.

**Physiology of flowering & Fruit Development:** Role of Photoperiodism and Vernalization in flowering, Genetic and molecular analysis of flower development: ABC model; fruit development and ripening as regulated by hormones, Seed formation and deposition of stored reserves during seed development, Embryo maturation and desiccation, Germination, Physiology of Bud and seed dormancy.

**Programmed Cell Death (PCD), Senescence:** Types of cell death observed in plants, overview of senescence, Pigment metabolism, protein metabolism & nucleic acid degradation during senescence, impact of senescence on photosynthesis & oxidative

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Suggested Readings


Elective Practical Lab BOT E13: Advanced Plant Physiology

Suggested Laboratory Exercises

1. Study of effect of PEG induced water stress on seed germination.
2. Effect of various Plant growth regulators on leaf discs (Normal & senescing)
3. Effect of Red and Infrared light on seed germination and study of photomorphogenesis.
4. Hormonal regulation of leaves and petal senescence.
5. To study the rhythmic movements of plants.
6. Study of changes in starch and protein content during seed development.
7. Study of Tropic movements in Plants.
8. Estimation of Total Chlorophyll in Healthy leaves and senescing leaves.

BOT E04: PPH: PHYTOCHEMISTRY AND HERBAL MEDICINE

Historical background & Present status- Scope of Medicinal Botany, Indigenous medical system, Bioprospecting, Indigenous Knowledge system, Ayurveda, Siddha, Unani, Homeopathy, Tibetan and Folklore system of medicine. Need to Preserve Knowledge system.
Phytochemical screening of crude drugs: Crude Drugs: Scope & Importance, Classification (Taxonomical, Morphological Chemical, Pharmacological); Cultivation, Collection & Processing of Crude Drugs.

Extraction and Characterization of Phytochemicals: Extraction, isolation, purification, characterization of following phytoconstituents such as Carbohydrates & Derived Products; Glycosides- Digoxin, Sennosides (Digitalis, Aloe, Dioscorea- Extraction Methods); Tannins(Hydrolysable & Condensed Types); Volatile Oils (Clove, Mentha- Extraction Methods); Alkaloids: Caffeine, Atropine, Ergometrine, Morphine; Flavonoids- Rutin, Quercetin; Terpenoids-Taxol, Pyrethin; Saponins- Glycyrrhizinic acid, Diosgenin.

Analysis of Phytochemicals: Standardization of phytopharmaceuticals by: UV, IR, HPLC, and HPTLC, GCMS techniques. Preliminary Screening, Assay of Drugs-Biological Evaluation/Assays, Microbiological Methods; Methods of Drug Evaluation (Morphological, Microscopic, Physical & Chemical); Processing, equipment and analytical profiles, Sterility, stability and preservation of extracts; Drug Adulteration - Types of Adulterants.

Indian Trade in Medicinal and Aromatic plants: Export potential of Indian medicinal herbs. Indian medicinal plants used in cosmetics and aromatherapy. Spices and their exports, Classification of medicinal plant based industry. Production and utilization of medicinal plants and their products in India. List of medicinal plants cultivated in India. Global regulatory status of herbal medicines: Patents: Indian and international patent laws, Recent amendments as applicable to herbal/natural products and processes.

Elective Practical Lab BOT E14: Phytochemistry and Herbal Medicine

Suggested Laboratory Exercises:

2. Demonstration of Soxhlet extraction method.
3. Solvent Screening for various phytochemical extractions.
4. Extraction methods of Phytochemicals (alkaloids, tannins, glycosides, Flavonoids)
5. Reagent test of alkaloids determination in plants (Dragendorff’s reagent, Meyer’s reagent, Hager’s reagent, Wagner’s Test).
6. Phytochemical screening and detection of carbohydrates (Molisch’s Test, Benedict’s test, Fehling’s Test)
7. Phytochemical screening and detection of glycosides, saponins, phytosterols, flavonoids, and tannins extracted from plant samples (at least one assay for each type of phytochemical).
8. Demonstration of counter current extraction, microwave-assisted extraction, ultrasound extraction (sonication), supercritical fluid extraction, and distillation techniques (water distillation, steam distillation, phytonic extraction (with hydro fluorocarbon solvents).

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Text/References Books:

6. Goodman and Gilman; Pharmacological Basis of Therapeutics, McGraw Hill
12. J.C. Willis: Pharmacognosy
13. C.K. Kokate: Pharmacognosy
25. Unani Pharmacopoeia.

Cluster: PE- Plant Ecology

BOT F01: PE: Ecosystem Ecology

Grassland Ecosystems - Characteristics of grasslands, stratification, grasslands and grazing, grasslands and drought, grassland and animal life, Grasslands types with special reference to Prairie and Savannah, ICAR classification of Indian grasslands, Significance of grasslands

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Suggested Readings

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**Elective Practical Lab BOT F11: Ecosystem Ecology**

**Suggested Laboratory Exercises**

- Find out stomatal index of Xerophytes (*Nerium, Calotropis, Zizyphus*) growing in your locality.
- Study of trichomes of xerophytes (*Zizyphus, Lantana, Calotropis, Aerva*) growing in your locality.
- Study spread of root system of a perennial species in the soil.
- Study ecological adaptations of halophytes in your nearby area.
- Seed Viability by T.T.C. method
- Dormancy (seed coat & temperature) in seeds.
- Soil moisture and temperature at different depths.
- Salinity of soil sample.
- Study of Canopy and Basal Cover of trees in your study area.
- Estimate primary productivity of a water body by light and dark bottle method.
- Mean leaf area of 2 plant Species growing in your area by graph method.
- Relative humidity by hair hygrometer.
- Light intensity by lux meter.
- Mark hot and cold desert on world map.

**BOT F02: PE: CONSERVATION BIOLOGY**

**Introduction:** Global as well as Indian Prospective - Biodiversity of World, India and Rajasthan. Hotspots or Important Biodiversity Area of India and Rajasthan, Biodiversity Categorization.


**Current issues in Biodiversity Loss:** Community based conservation approach, Impact of climate change on species diversity, Human-wildlife conflict, Illegal trading (special reference to medicinal plants and timber), Habitat loss and Habitat fragmentation.

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Elective Practical Lab BOT F12: Conservation Biology

Suggested Laboratory Exercises:

1. Visit any protected area (National park/Sanctuary) and document red listed species.
2. Biodiversity assessment in your area.
3. Documentation of wildlife protection act for the plants species.
5. Documentation of IUCN and CITES species of Rajasthan plants.
6. Visit any Sacred groves and documentation of their species.
7. Visit any tribal community and describe their role for conservation.
8. In situ conservation of germplasm/plant species
9. Preparation of list of endangered plant species in your area
10. Analysis and discussion of data of Project Tiger, Project Elephant, Project Rhino and GIB

Books Recommended:


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BOT F03: PE: Environmental Biology

Air Pollution: Important Primary (CO, CO2, Oxides of Sulphur& Nitrogen, H2S, Chlorine, Particulates, Odour Producing compounds) & Secondary Air Pollutants (Smog, Acid rain, Primary Photochemical reaction, Formation of ozone and peroxyacetyl nitrate in air), Effects of air pollutants on Buildings & Monuments, plants, man and animals; Biomonitoring, Air pollution control (particulates and gaseous pollutants), Green belt, Ozone depletion, mechanism of depletion, control strategies; 12

Water Pollution: Eutrophication- Process and Control; Oil Pollution, Thermal Pollution, Heavy metal Pollution, Treatment, Disposal & Recycling of Wastewaters, drinking water standards, Minimum National Standards 10

Solid & Hazardous waste management & Resource Recovery: Solid wastes, Types, collection, Shrinking waste streams: 4Rs (Refuse, Reduction, Recycle & Reuse), composting, energy from waste, demanufacturing; Methods of disposal: Land fill, Open dumps, Exporting waste; Hazardous waste: Definition, disposal and management 08


Climate Issues: Greenhouse gases (CO2, CH4, N2O, CFCs: sources, trends and role) and consequence of greenhouse effects (CO2 fertilization, global warming, sea level rise, Biodiversity erosion), Carbon footprints, Carbon sequestration, Applications of GIS and Remote Sensing technology in environmental studies, the future of planet earth. 10


Environmental concerns: Environment auditing, Ecological footprints, Environment Impact Assessment, Bioindicator and biomarkers of environmental health; Environmental economics, Ecopolitics and green policies; Ecolabel, Rain water harvesting, Orans, Indira Gandhi Canal and its, ecological implication, water logging & salinity problems- The management alternatives. 10

Suggested Readings


• I.P.Abrolamd V.V. Dhruvanarayana (Editors) 1990. Technologies for Wasteland Development. ICAR, New Delhi.

• J.S. Singh, K.P. Singh, Shuda Gupta: Environmental Biology


**Elective Practical Lab BOT F13: Environmental Biology**

**Suggested Laboratory Exercises**

• To estimate pH, EC and Secchi Disc transparency for polluted and unpolluted water bodies.

• To estimate Chemical Oxygen Demand of polluted water sample.

• To estimate Biological Oxygen Demand of polluted water sample.

• To estimate inorganic phosphorus content in water samples collected from polluted and unpolluted water bodies.

• To estimate Total hardness, calcium and magnesium content in water samples collected from polluted and unpolluted water bodies.

• To estimate chloride content in water samples collected from polluted and unpolluted water bodies.

• To estimate Total alkalinity in water samples collected from polluted and unpolluted water bodies.

• To determine diversity indices (Shannon-Wiener, concentration of dominance, species richness, equitability and β-diversity) for polluted and unpolluted water bodies.

• Chlorophyll content of plant species growing in polluted and unpolluted habitat

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Cluster: GE- Plant Genetic Engineering

BOT G01: GE: GENOMICS & PROTEOMICS


Approaches to analyze differential expression of genes - ESTs, SAGE, microarrays and their applications, gene tagging, gene and promoter trapping, knockout and knock-down mutants. Transcriptome, Transcriptomics, RNA interference and gene silencing, genome imprinting, small RNAs-biogenesis and functions, role of small RNAs in heterochromatin formation and gene silencing, tools to study methylome and histone modifications.


Tools and techniques of proteome analysis: 1-D and 2-D gel electrophoresis, DIGE (Differential In Gel Electrophoresis), Image analysis of 2D and DIGE gels; spot detection & quantitation, gel matching, data analysis and presentation, Liquid Chromatography and Multidimensional Chromatography; Protein Identification by Mass Spectroscopy (MALDI/TOF), LC/MS-MS for identification of proteins, Peptide Mass Fingerprinting, Protein de novo sequencing.

Analysis of proteins by different biochemical and biophysical procedures: CD (Circular Dichroism), NMR, Analysis of post-translational modifications and protein-protein interactions; protein chips and arrays, Proteome Databases: Protein Sequence Database, SWISS-PROT, PROSITE, PDB etc. other protein related bioinformatics tools (ExpASy, PFAM). Applications of proteomics in medicine, toxicology and Pharmaceuticals.

Elective Practical Lab BOT G11: Genomics & Proteomics

Suggested Practical Exercises:

1. Demonstration and listing of sequence retrieval online tools.
2. Demonstration and listing of sequence submission online tools.
3. Listing and demonstration of Protein and DNA Sequence Databases and their utilities.
4. Demonstration of DNA and Protein Array Technology and applications.
5. Reverse transcription-PCR to examine gene expression.
6. Real-time PCR to quantify gene expression.
7. Northern and Western Blotting analysis.
8. Demonstration of Instrumentation (MALDI/TOF, LC-MS-MS, 2DGE) by visit or audio-visual medium.
9. Protein separation techniques (Chromatography-Ion-Exchange, Gel Filtration, Affinity, Ultrafiltration, Recombinant protein separation techniques).
10. Comparison of Next-generation sequencing methods (by Chart/poster preparation).
11. Any other exercises designed by course teacher as per the syllabus.

SUGGESTED READINGS:


BOT G02: GE: ADVANCE GENETIC ENGINEERING
AND MOLECULAR PHARMING

Enzymes used in genetic engineering: Restriction nucleases: exo&endo nucleases, DNA modifying enzymes and their mechanisms of action: DNA polymerases, DNAs, RNAes, Reverse transcriptase, Polynucleotide phosphorylase, Polynucleotide kinase, Alkaline phosphatase, Ligase, Terminaldeoxynucleotidyltransferase.
Cloning and expression hosts: Characteristics of cloning and expression host, bacterial, yeast, plant and mammalian host systems for cloning and expression of genes.

Vectors for plant transformation: Basic features of vectors (Promoters, terminators & sequences influencing gene expression, selectable markers & reporter genes, origin of replication), Vectors for cloning - Plasmids, Bacteriophage λ vectors, Cosmids, BAC and YAC vectors, Shuttle vectors, Expression vectors, Co-integrative and binary vectors for plant transformation

Heterologous gene expression in plants and Genetic manipulation of plants for Herbicide tolerance (Roundup ready Soybean), Insect resistance (Bt Cotton), stress tolerance, disease resistance, Improvement of crop yield and quality (Post-harvest loses, longer shelf-life of fruits (flavrsvavr tomato), color manipulation of flowers (Blue rose), Making of Golden Rice Genome editing (TALEN & CRISPR) and its applications in crop improvement. Applications of DNA based molecular markers (RFLP, RAPD and AFLP) in plant biotechnology.

Science and society: Public acceptance of genetically modified crops (Public concerns, current status of transgenic crops, regulation of GM crops, Cisgenic crops, and products), Introduction to Intellectual property, Biosafety guidelines, Environmental release of GMO’s, Risk analysis, Risk Assessment and Risk management.

Molecular Pharming in Plants
Introduction and brief history of plant molecular farming; Unique properties of host species for molecular farming (tobacco, alfalfa, white clover, lettuce, spinach, dry seed crops, oil crops, fruit and vegetable crops); Types of plant expression systems for molecular pharming (Stable nuclear transformation, plastid transformation system, virus-infected plants, transiently transformed leaves, hydroponic cultures, hairy roots, shooty teratomas, suspension cell cultures, Oleosin system); Comparison, advantage and disadvantages of production system of molecular pharming (bacteria, yeast, transgenic animal, plant cell culture and transgenic plants).

Plant Made Pharmaceuticals (PMPs)
(1) Plant Made Pharmaceutical and Therapeutic proteins (Human Serum Albumin, Human Insulin-like Growth Factors, Human Interferon, Anti-Microbial Peptides, Therapeutic Enzymes, Lactoferrins)* (2) Industrial proteins (3) Monoclonal recombinant antibodies (4) Antigens (Edible vaccines- Rabies, Hepatitis B, Respiratory Syncytial Virus, Enterotoxic E.coli and Vibrio cholerae). Detailed account of Biopharmaceuticals already in market and close to market.
Elective Practical Lab BOT G12: Advance Genetic Engineering and Molecular Pharming

Suggested Laboratory Exercises:-

1. Aseptic culture techniques for establishment and maintenance of cultures for genetic engineering.
2. Preparation of suitable media to grow transformed E. coli cells.
3. Activity assay of various enzymes involved in genetic engineering.
4. Analysis of different Plant DNA samples by molecular markers.
5. Agrobacterium culture, selection of transformants, reporter gene (GUS) assays.
7. PCR and Gel electrophoresis for gene cloning.
8. Demonstration of GLPs and Biosafety in research labs.
10. Transformation of competent cells of E.coli with plant transformation vector.
11. Mobilization of Ti plasmid from common laboratory host (E. coli) to an Agrobacterium tumefaciens strain.
13. Assignments on topics such as GMO, genome editing, PCR, gene cloning and molecular farming etc.

Suggested Readings:

3. Molecular Farming in Plants: Recent Advances and Future Prospects by Aiming Wang (Editor), Shengwu Ma (Editor), Springer, 2011.
Cluster: GEN- GENERAL

BOT H01: GEN: PRINCIPLES OF PLANT BREEDING

Overview & Historical perspectives: History of Plant Breeding-the pioneers, their theories and plant breeding techniques.

Population and quantitative Genetic principles: Concept of Population, gene pool, gene frequency and inbreeding and its implications in breeding. Qualitative genetics versus Quantitative genetics, the concept of Population Improvement.

Reproductive systems: Importance of Mode of Reproduction, Types of Reproduction, Autogamy, Haploids and double haploids: their application in plant breeding, Allogamy, Inbreeding depression, hybrid vigour, Hybridization, wide crosses, clonal propagation and In vitro culture.

Germplasm for Breeding: Variation-Types, origin and scale, Plant Domestication- Centres & Models, Plant Genetic resources-Importance & Sources of Germplasm. Concept of Gene pools, Crop vulnerability, Germplasm conservation: In situ & Ex situ, Types of Germplasm collection, Germplasm storage technologies, Plant explorations & Introductions & their impact on agriculture.

Breeding Objectives: Yield and morphological trait- Yield potential, Harvest Index, breeding for lodging resistance, shattering resistance, plant stature & early maturity, Quality traits- breeding for improved protein content, improved fatty acid content, seedlessness in fruits, delayed ripening & novel traits, Breeding for resistance to disease & insect pests – Resistance Breeding strategies; Abiotic Stresses – Breeding for drought resistance, cold tolerance, salt tolerance, heat stress, aluminium toxicity, oxidative stress, resistance to water logging.


Marketing and Societal issues in Breeding: Performance and Evaluation for crop cultivar release, Seed certification and commercial seed release, Regulatory and Legal issues. Value driven concepts and social concerns, International Plant breeding efforts, Plant cultivar...
Elective Practical Lab BOT H11: Principles of Plant Breeding

Suggested Laboratory Exercises:

1. Vegetative propagation methods of important crops of the locality.
2. Emasculating, selfing and crossing techniques.
3. Floral biology in self pollinated species
4. Floral biology in cross pollinated species
5. Floral biology in self pollinated species
6. Selection methods in segregating populations and evaluation of breeding material
7. Germplasm conservation methods- In situ & Ex situ methods
8. Haploid production.
9. In situ & ex situ conservation methods
10. Analysis of variance (ANOVA).
11. Maintenance of experimental records
12. Field inspection at different growth stages to study various breeding objectives.

Suggested Readings:

BOT H02: GEN: BIOINFORMATICS AND BIOSTATISTICS

Concepts of Bioinformatics: Introduction and future prospects; Applications in genomics and proteomics; Public databases; Gene bank; Database searches: sequence retrieval systems; Similarity searching: BLAST, FASTA; Multiple sequence alignment: CLUSTALW; Detecting functional sites in DNA; Motif and domain prediction and analysis; Identification of open reading frames (ORF); Gene annotation technology.

Databases and online tools: Biological Databases: - Types and applications; Sequence databases: - GenBank, EMBL, DDBJ, PIR-PSD, SWISS-PROT, Structure Databases: - PDB, SCOP, NDB; Derived Databases: - PROSITE, PRINTS, TIGR, Online tools: - Genetool, STRING; I-TASSER, Bioedit, BioGRID; MEGA; Sequin, Bankit

Applications of Bioinformatics: Computational methods for sequence analysis; Dot blot and dynamic programming methods; Phylogenetic analysis; Virtual and electronic cell; Internet tools for DNA sequence translation; Restriction enzyme mapping; Prediction of secondary structure of proteins; Application tools: primer designing, molecular mapping and concept and tools of computer aided drug designing

Fundamentals of statistics: Arithmetic mean, median, mode: theory and simple numerical problem; Measures of variation: standard deviation, variance, coefficient of variation; Correlation: types and methods: simple, multiple, linear and nonlinear correlation; Spearman's correlation, rank correlation; Regression: linear and curvilinear regression (for two variable X and Y only), Regression lines by least square method; regression equations of X on Y and Y on X only; Sample size; Power of study.

Tests of significance: Null hypothesis; Standard error, Level of significance; Degrees of freedom; Significance of mean for large samples; Significance in means for small samples (students t-test); Significance in ratio of two samples; F test (for difference between variance of two samples); Chi square test; Analysis of variance test (ANOVA) for one and two way classification; Calculation of an unknown variable using regression equation.

Laws of probability, theorem of total probability
Elective Practical Lab BOT H12: Bioinformatics and Biostatistics

Suggested Laboratory Exercises

1. Introduction to bioinformatics databases (any three): NCBI/PDB/DDBJ, Uniprot, PDB etc.
2. Sequence retrieval using BLAST
3. Sequence alignment
4. Phylogenetic analysis using clustalW
5. Protein structure prediction
6. Picking out a given gene from genomes using Genscan or other softwares (promoter region identification, repeat in genome, ORF prediction).
7. Prediction of different features of a functional gene
8. Determination of Statistical averages/ central tendencies
   a) Arithmetic mean b) Median c) Mode
9. Determination of measures of Dispersion a) Mean deviation b) Standard deviation and coefficient of variation c) Quartile deviation
10. Tests of Significance-Application of following a) Chi- Square test b) t- test c) Standard error
11. To learn graphical representations of statistical data with the help of computers (e.g. MS Excel).

Suggested Readings

1. Introduction to Bioinformatics, Arthur M. Lesk, Oxford University Press.
2. Introduction to Bioinformatics, Attwood, Pearson Education.
8. Bioinformatics - from Genomes to drug. 2 volumes by Lengenar.
10. Bioinformatics: Sequence and Genome analysis
11. Introduction to Bioinformatics by Altwood.
BOT H03: GEN: PLANT RESOURCE UTILIZATION

Economic importance of microbes, with special reference to Bacteria and algae of your locality and found in extreme habitats 3

Role of fungi in industries with reference to the production of alcohol, organic acids, antibiotics and enzymes 3

Plant Biodiversity: Concept, status in India, utilization and concerns. Sustainable development: Basic Concepts. Origin of agriculture 6

World centers of primary diversity of domesticated plants. The Indo-Burmese centre, plant introductions and secondary centers. 3

Origin, evolution, botany, cultivation and uses of (i) Food, forage and fodder crops, (ii) ornamental plants, plants used in sericulture, as petro crops, in narcotics, as mastigatories, fumitories and fiber crops, (iii) medicinal and aromatic plants and (iv) vegetables, fruits, spices condiments & oil-yielding crops. 10

Important fire-wood and timber-yielding plants and non-wood forest products (NWFPs) such as bamboos and rattans, Raw materials for paper making, gums, tannins, dyes and resins 5

Green revolution: Benefits and adverse consequences. Recent trends of research for meeting world food demands. Plants used as avenue trees for shade, pollution control and aesthetics. 4

General account of the activities of Botanical Survey of India (BSI), National Bureau of plant Genetic Resources (NBPGR), Indian Council of Agricultural Research (ICAR), Council of Scientific and Industrial Research (CSIR), and the Department of Biotechnology (DBT) for conservation and non-formal conservation efforts.

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Basic Horticultural techniques (soil preparation, bed preparation transplantation and pruning, vegetative cultivation of cut flowers (Roses, Gerberas and Carnations), Bonsai- Types, forms, structure and styles, Green House technique: Importance, types and operation techniques. Soil less culture (hydroponics concept)

Suggested Reading:-

1. Introduction to Agricultural Botany in India Chalam, 1965: G.V.I. Venkateshwarlu J.; Asia Publishing House, New Delhi.
4. Contributions of Indian Ethnobotany : S.K. Jain


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Elective Practical Lab BOT H13: Plant Resource Utilization

Suggested Laboratory Exercises

The Practical course is divided into three units: (1) Laboratory work, (2) Field survey and (3) Scientific visits.

Laboratory Work

1. Food crops: Wheat, rice, maize, chickpea (Bengal gram), potato, sweet potato, sugarcane. Morphology, anatomy, biochemical tests for stored food materials.

2. Forage/fodder crops: Study of any five important crops of the locality (for example fodder sorghum, bajra, berseem, khejari, Ardu, zizyphus.

3. Plant fibers:
   (a) Textile fibres: cotton, jute, linen, sunn hemp, Cannabis.
   (b) Cordage fibres: coir (c) Fibres for stuffing: silk cotton or kapok

Morphology, anatomy, (microscopic) study of whole fibres using appropriate staining procedures.

4. Medicinal and aromatic plants: Depending on the geographical location college/university select five medicinal and aromatic plants each from a garden crop field (or from the wild only if they are abundantly available).

5. Vegetable oils: Mustard, groundnut, soybean, coconut, sunflower, castor. Morphology, microscopic structure of the oil-yielding tissues, tests for oil and iodine number.

6. Gums, resins, tannins, dyes: Perform simple tests for gums and resins. Prepare a water extract of vegetable tannins (Acacia, Terminalia, tea, Cassia spp., myrobalans) and dyes (turmeric, Indigo, Buteamonomosperma, Lawsoniainermis) and perform tests to understand their chemical nature.

7. Firewood and timber yielding plants and NWF's

Field Surveys
a. Prepare a short list of 10 most important sources of firewood and timber in your locality. Give their local names, scientific names, and families to which they belong. Mention their properties.
b. Prepare an inventory of the bamboos and rattans of your area giving their scientific and local names and their various uses with appropriate illustrations.
c. A survey of a part of the town or city should be carried out by the entire class, in batches. Individual students will select one avenue/road and locate the trees planted on a graph paper. They will identify the trees, mention their size, canopy shape, blossoming and fruiting period and their status (healthy, diseased, infested, mutilated, misused or dying) and report whether or not the conditions in which they are surviving are satisfactory. The individual reports will be combined to prepare a larger map of the area, which can be used for subsequent monitoring either by the next batch of students/teachers/local communities/NGOs or civic authorities. The purpose of exercise in item C above is to make the students aware of the kinds of trees and value in urban ecosystems and ecological services.

Scientific Visits*
The students should be taken to one of the following:
i. A protected area (biosphere reserve, national park, or a sanctuary)
ii. A wetland
iii. A mangrove
iv. National Bureau of Plant Genetic Resources, New Delhi-110012 or one of its field stations.
v. Head Quarters of the Botanical Survey of India or one of its Regional Circles.
vi. A CSIR Laboratory doing research on plants and their utilization.
vii. An ICAR Research Institute or a field station dealing with one major crop or crops.
viii. A recognized botanical garden or a museum (such as those at the Forest Research Institute, dehradun, National Botanical Institute, Lucknow, Tropical Botanical Garden and Research Institute, Trivendram), which has collection of plant products. Note: The students are expected to prepare a brief illustrated narrative of the field survey and scientific Visits.
After evaluation, the grades awarded to the students by the teachers should be added to the field assessment of the practical examination.

**BOT H04: GEN: METHODS IN PLANT SCIENCES**

**Centrifugation:** Plant Extraction and isolation of cell organelles, Basic principle, RCM, classification, Ultracentrifuge, Precautions and applications.  

05


08

**Immunotechniques:** Antibody generation, Detection of molecules using ELISA, RIA, western blot, immunoprecipitation, flow cytometry and immunofluorescence microscopy, detection of molecules in living cells, in situ localization by techniques such as FISH and GISH.  

07

**Biophysical Method:** Molecular analysis using UV/visible, fluorescence, circular dichroism, NMR and ESR spectroscopy Molecular structure determination using X-ray diffraction and NMR, Molecular analysis using light scattering, different types of mass spectrometry.  

08

**Radiolabeling techniques:** Detection and measurement of different types of radioisotopes normally used in biology, incorporation of radioisotopes in biological tissues and cells, molecular imaging of radioactive material, Autoradiography-Principle, method, use in gene probe molecules, safety guidelines.  

06

**Microscopic techniques:** Visualization of cells and sub cellular components by light microscopy, resolving powers of different microscopes, microscopy of living cells, scanning and transmission microscopes, different fixation and staining techniques for EM, freeze-etch and freeze-fracture methods for EM, image processing methods in microscopy.  

06

**Statistical Methods:** Measures of central tendency and dispersal; probability distributions (Binomial, Poisson and normal); Sampling distribution; Difference between parametric and non-parametric statistics; Confidence Interval; Errors; Levels of significance; Regression and Correlation; t-test; F-test; Analysis of variance; X2 test.  

10

**Bioinformatics (Definition, history, applications & scope):** Biological databases and Sequence analysis, BLAST and FASTA, Multiple sequence alignment, Phylogenetic analysis and Bioinformatics in Drug Discovery.  

08

Dy. Registrar  
(Academic)  
University of Rajasthan  
JAIPUR
Elective Practical Lab BOT H14: Methods in Plant Sciences

Suggested Laboratory Exercises:

1. Instrumentation in Lab.
2. Microscopy- SEM, TEM, Florescence, Staining & fixation procedures
3. Centrifugation technique for isolation of cell organelles
5. Measurement of cell size using stage micrometer and ocular micrometer.
6. Quantification of DNA by UV-Visible spectrophotometer.
7. Quantification of RNA by UV-Visible spectrophotometer
8. NMR spectroscopy technique.
9. IR spectroscopy technique.
10. Numericals related to biostatistics
11. ELISA, RIA, Flow cytometry
12. BLAST and FASTA
13. Phylogenetic analysis
14. Any other Exercise within scope of the syllabus

Suggested Readings:


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